



describe the synthesis of an unsymmetrical bis-lexitropsin–CBI conjugate, which contains two different lexitropsins. In our previous work,<sup>6</sup> the CBI moiety was obtained by an *in situ* primary radical trap with TEMPO. Here, the CBI moiety was synthesized by using a more concise and shorter route which was recently developed by Patel and co-workers.<sup>8</sup>

Deprotonation of carbamate **1**<sup>6</sup> using NaH, followed by alkylation of the resulting anion with 1,3-dichloropropene in the presence of the phase transfer catalyst Bu<sub>4</sub>N<sup>+</sup>I<sup>-</sup> gave a mixture of *Z* and *E* isomers of vinyl chloride **2**. Selective reduction of the nitro group of **2** using hydrazine,<sup>9</sup> followed by protection of the amino group, provided **3**, the desired precursor for the intramolecular aryl radical cyclization on to a tethered vinyl chloride.<sup>8</sup> A deoxygenated solution of **3** in dry benzene was heated at reflux for 15 h in the presence of 2 equiv. of Bu<sub>3</sub>SnH and a catalytic amount of AIBN to give the fully protected bifunctionalized CBI prodrug form, racemic **4**.<sup>†</sup> Although not investigated in detail, no reaction occurred when nitro compound **2** was treated under the same conditions as amine **3**.

Detachment<sup>10</sup> of the Fmoc group from **4**, followed by coupling with polypyrrole carboxamide **5**<sup>11</sup> using HOBt and EDCI as the coupling agents<sup>11b,c</sup> afforded the hybrid **6**. Hydrogenolysis<sup>8a</sup> of **6** served to remove the benzyl ether almost quantitatively and provided **7**. Acid-mediated deprotection of **7**, followed by coupling with polyimidazole carboxamide **8**<sup>11</sup> using EDCI provided the final bis-lexitropsin–CBI precursor conjugate **9**<sup>‡</sup> in fair yield.

In summary, we have described a synthesis of the bis-functionalized CBI precursor containing two different protective groups and obtained the corresponding unsymmetrical bis-lexitropsin conjugate. Results on the DNA sequence preferences and biological evaluation will be reported in due course.

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## Notes and references

<sup>†</sup> Selected data for **4**:  $\delta_{\text{H}}$ (360 MHz, acetone-*d*<sub>6</sub>) 9.02 (s, NH), 8.38 (s, 1H, C6-H), 7.88–7.30 (m, 16H, Ar-H), 5.30 (s, 2H, PhCH<sub>2</sub>O), 4.50 (d, 2H, *J* 6.9, CH<sub>2</sub> in Fmoc), 4.30 (t, 1H, *J* 6.9, CH in Fmoc), 4.22–4.05 (m, 3H, C1-H, C2-H), 4.01 (dd, 1H, *J* 3.1, 11.1, CHHCl), 3.70 (dd, 1H, *J* 8.4, 11.0, CHHCl), 1.58 (s, 9H, Boc-H); Calc. for C<sub>40</sub>H<sub>37</sub>N<sub>2</sub>O<sub>5</sub>Cl: C, 72.66; H, 5.64; N, 4.24. Found C, 72.55; H, 5.74; N, 4.20%.

<sup>‡</sup> Selected data for **9**:  $\delta_{\text{H}}$ (360 MHz, DMSO-*d*<sub>6</sub>) 10.35 (s, 1H), 10.30 (s, 1H), 10.18 (s, 1H), 9.90 (s, 1H), 9.86 (s, 1H), 9.75 (s, 1H), 9.62 (s, 1H), 9.59 (s, 1H), 8.55 (d, 1H, *J* 2.0, C6-H), 7.97 (s, 1H, C4-H), 7.86 (dd, 1H, *J* 2.0, 7.5, C8-H), 7.74 (d, 1H, *J* 7.5, C9-H), 7.64 (s, 1H, Im-H), 7.60 (s, 1H, Im-H), 7.51 (s, 1H, Im-H), 3.31 (d, 1H, *J* 1.5, Py-H), 7.24 (d, 1H, *J* 1.5, Py-H), 7.21 (d, 1H, *J* 1.5, Py-H), 7.16 (d, 1H, *J* 1.5, Py-H), 7.07 (d, 1H, *J* 1.5, Py-H), 6.88 (d, 1H, *J* 1.5, Py-H), 4.7–3.6 (m, 23H, NCH<sub>3</sub>, ClCH<sub>2</sub>, C1-H, C2-H), 2.28 (t, 2H, *J* 7.3, COCH<sub>2</sub>), 2.20 (t, 2H, *J* 7.4, COCH<sub>2</sub>), 1.63–1.53 (m, 4H, COCH<sub>2</sub>CH<sub>2</sub>), 0.91–0.86 (m, 6H, COCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); Calc. for C<sub>54</sub>H<sub>58</sub>N<sub>17</sub>O<sub>9</sub>Cl: C, 57.15; N, 21.17. Found C, 57.06; N, 20.97%.

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